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PATENT APPLICATION FOR

**COMPOSITION AND METHOD FOR PREVENTING AND TREATING  
SINUSOIDAL OBSTRUCTION SYNDROME AND RADIATION-INDUCED  
LIVER DISEASE**

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## **TITLE OF INVENTION**

Composition and Method for Preventing and Treating Sinusoidal Obstruction Syndrome and Radiation-Induced Liver Disease

## **CROSS-REFERENCE TO RELATED APPLICATIONS**

This application claims the benefit of U.S. Provisional Application No. 60/271,780, filed February 27, 2001, the entire disclosure of which is hereby incorporated by reference in its entirety for all purposes.

## **FIELD OF THE INVENTION**

This invention relates to the use of matrix metalloproteinase (MMP) inhibitors in the prevention and treatment of Sinusoidal Obstruction Syndrome; in particular the present invention relates to the prevention and treatment of chemotherapy- and radiation-induced liver complications.

## **STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH**

This invention was made with government support under NIDDK, Grant DK46357 awarded by the National Institutes of Health.

## **BACKGROUND OF THE INVENTION**

The present invention is directed to the use of matrix metalloproteinase ("MMP") inhibitors to prevent and treat Sinusoidal Obstruction Syndrome ("SOS"). SOS, also known as hepatic venoocclusive disease, was first diagnosed in cases of liver disease caused by the ingestion of herbal teas or food sources containing pyrrolizidine alkaloids from *Crotalaria*, *Heliotropium* and *Senecio* or from the consumption of bread made from inadequately winnowed wheat contaminated by seeds from these plants. With the modern development of chemotherapy, cases of SOS developed from the long-term use of azathioprine for immunosuppression after renal and liver transplantation and from the use of chemotherapeutic agents. Liver complications of chemotherapy are seen most commonly after high dose chemotherapy, with or without total body irradiation, or high dose radiation to the liver. Liver toxicity is not an uncommon side effect of high-dose chemotherapy. Liver toxicity also occurs after chemotherapy and/or liver irradiation

when there is no bone marrow transplantation and hence, conditioning regimens used for marrow ablation are the most common cause of SOS.

SOS is a common complication of chemotherapy with gemtuzumab ozogamicin<sup>2</sup> or actinomycin D,<sup>3</sup> or after long-term immunosuppression with azathioprine in kidney or liver transplantation patients. Other chemotherapeutic agents associated with liver toxicity and SOS include dacarbazine, cytosine arabinoside, mithramycin, 6-thioguanine, urethane, indicine N-oxide, alone and in combination. Milder forms of liver disease from chemotherapy which share the key aspect of sinusoidal endothelial cell injury include nodular regenerative hyperplasia, sinusoidal dilatation and peliosis hepatis. Combinations of irradiation and chemotherapy have also led to the development of SOS. For example, treating nephroblastoma (Wilms' tumor) with dactinomycin and abdominal irradiation has led to SOS.

Radiation-induced liver disease is a condition that shares some of the features of SOS, although there are differences in clinical presentation, histology and time course. Radiation-induced liver disease is seen with radiation doses in excess of 30 to 35 Gy in adults.

SOS has significant morbidity and mortality. The severity of SOS can be classified as mild (SOS is clinically obvious, but requires no treatment and resolves completely), moderate (SOS that causes signs and symptoms requiring treatment such as diuretics or pain medications, but resolves completely) or severe (SOS that requires treatment but that does not resolve before death or day 100.<sup>15, 16, 17, 22</sup> Some patients have subclinical liver damage, evinced by histologic evidence of liver toxicity in the absence of clinical signs and symptoms.<sup>18</sup> Despite deep jaundice, patients with severe SOS seldom die of liver failure, but rather from renal and cardiopulmonary failure.<sup>15, 16, 23, 24</sup>

A clinically useful model for predicting the outcome of SOS after cyclophosphamide-based regimes is derived from rates of increase of both bilirubin and weight in the first two weeks following transplantation.<sup>25</sup> Furthermore, a poor prognosis correlates with higher serum AST and ALT values, higher wedged hepatic venous pressure gradient, development of portal vein thrombosis, doubling of the baseline serum creatinine, and decreasing oxygen saturation.<sup>19, 20, 21, 14, 26</sup> There is currently no prophylactic treatment for either SOS or radiation-induced liver disease, and there are no proven

therapeutic remedies with high efficacy. The only therapeutic modality with some proven efficacy is the combination of heparin plus tissue plasminogen activator. However, this combination can only be safely used in a very limited group of patients and has efficacy in less than 30% of this limited population of patients.

SOS is the dose-limiting toxicity for several chemotherapeutic drugs and limits patient eligibility. A prophylactic treatment of SOS would have a significant impact on the ability to use high dose chemotherapy. Development of therapies to treat SOS after onset of the disease would be of value in unexpected cases of chemotherapy-induced liver disease.

The molecular events have been best characterized in the rat monocrotaline model. Monocrotaline, the pyrrolizidine alkaloid found in *Crotalaria*, is one of the best-studied toxins involving SOS.<sup>5,6,4</sup> The monocrotaline model of SOS has the same histologic characteristics as the human disease, as well as the same "clinical features," i.e., hyperbilirubinemia, hepatomegaly, and ascites formation. In this model, the first morphologic change noted by electron microscopy is loss of the sinusoidal endothelial cell fenestration and the appearance of gaps in the sinusoidal endothelial cell barrier.<sup>8</sup> Studies with in vivo microscopy and confirmation by electron microscopy have shown that sinusoidal endothelial cells round up, and red blood cells begin to penetrate into the space of Disse beneath the rounded up endothelial cells and dissect off the sinusoidal lining. The sloughed sinusoidal lining cells (i.e., Kupffer cells, sinusoidal endothelial cells, and stellate cells) embolize downstream and obstruct sinusoidal flow.<sup>7</sup> By the time hepatocyte necrosis is observed, there is extensive denudation of the sinusoidal lining. Early on, there is loss of Kupffer cells, but there is a significant influx of monocytes within the sinusoids, which exacerbates the obstruction of sinusoidal flow by the embolized sinusoidal lining cells. The rounding up or swelling of sinusoidal endothelial cells is the initiating event in SOS and leads to dissection of the sinusoidal lining, which embolizes and blocks the microcirculation.

The initial change to the sinusoidal endothelial cell is morphologically similar to the change in these cells in cold preservation injury. In studying cold preservation cells, Strasberg et al. showed that the upregulation of matrix metalloproteinases ("MMPs") is involved in changes to the sinusoidal cell.<sup>11</sup> Prior to the present invention, it was

unknown in the art whether MMPs may be involved in SOS. The inventors of the present invention discovered the relationship between MMPs and the development of SOS. With the discovery of the mechanism that initiates SOS, the inventors were able to develop therapies for the prevention and treatment of SOS and radiation-induced liver disease.

## **SUMMARY OF THE INVENTION**

The present invention relates in general to the use of MMP inhibitors in the prevention and treatment of liver disease. Accordingly, the present invention provides means to prevent and treat SOS and radiation-induced liver disease.

In a first aspect of the invention, a method is provided for preventing and treating SOS.

In another aspect of the invention, a method is provided for preventing and treating liver complications of chemotherapy, including SOS, nodular regenerative hyperplasia, peliosis hepatis, immunosuppression-induced hepatic venoocclusive disease, and sinusoidal dilatation. It is also an objective of this invention to provide a means to prophylactically treat radiation-induced liver disease.

It is another object of this invention to provide a means to increase patient eligibility for multiple chemotherapeutic drugs by preventing SOS.

## **BRIEF DESCRIPTION OF THE DRAWING**

### **Figure 1. Prevention of SOS by MMP Inhibition**

This figure describes the effect of MMP inhibition in the in monocrotaline-induced model of SOS. Injury is rated as absent (-) or as one, two or three plus. The overall score reflects central vein (CV) endothelial damage, hemorrhage and coagulative necrosis: 2-3 points is considered mild SOS, 4-6 points is considered moderate SOS and 7-9 points is severe disease. The MMP2/MMP9 inhibitor used is 2-[(4-biphenyl)sulfonyl]amino]-3-phenyl-propionic acid.

As can be seen, on day four, monocrotaline induces severe SOS. This is completely prevented by 2-[(4-biphenyl)sulfonyl]amino]-3-phenyl-propionic acid and by doxycycline. On the other hand, the two chemically modified tetracyclines, anhydrotetracycline and isochlorotetracycline, which are doxycycline analogues that are weak MMP inhibitors, do not prevent SOS.

## DETAILED DESCRIPTION OF THE INVENTION

All scientific terms are to be given their ordinary meanings as understood by those of skill in the art, unless an alternate meaning is set forth below. In case of conflict, the definitions set forth in this specification shall control.

In the present invention, the term "Sinusoidal Obstruction Syndrome" or "SOS" is synonymous with the term "hepatic venoocclusive disease."

In the present invention, it is disclosed that an explanation for the rounding up of sinusoidal endothelial cells may be due to increased activity of MMPs. Because MMPs degrade extracellular matrix, increased MMP activity on the abluminal side of the sinusoidal endothelial cell would allow the cells to let loose from the space of Disse. In the experimental model, de novo synthesis of MMP-9 (gelatinase B) and increased MMP-9 activity occur 12 hours after monocrotaline, which coincides with rounding up of the sinusoidal endothelial cells.<sup>12</sup> Inhibition of MMP activity completely prevents SOS. MMP expression and activity are regulated by redox status and can be suppressed by glutathione and N-acetylcysteine.<sup>27-30</sup> Thus, the protective effect of glutathione and N-acetylcysteine appears to be due to inhibition of MMP activity.

The present invention extrapolates from the above model to disclose a method for using matrix metalloproteinase (MMP) inhibitors to prevent or treat SOS and radiation-induced liver disease. The present invention discloses that doxycycline, an MMP inhibitor, completely prevented SOS in a rat model and with human subjects. Further experimentation showed that this was a class effect as the MMP-2/MMP-9 inhibitor, 2-[(4-biphenylsulfonyl)amino]-3-phenyl-propionic acid (BPP) also completely inhibits SOS. A number of other MMP inhibitors will also be effective in the prevention and treatment of SOS and radiation-induced liver disease. For example, Marimastat, Prinomastat and RS-130,830 are potent inhibitors of the MMPs that are increased in the monocrotaline model of SOS. CGS 27023A, Solimastat, BAY 12-9566, Ro 32-3555, BMS-272591, Ilomastat, D2163 are also MMP inhibitors that could be used in humans. Metastat, Neovastat, and Periostat also have potential therapeutic uses in treating and preventing SOS and radiation-induced liver disease.

As described in EXAMPLE I below, the protective effect of both doxycycline and of 2-[(4-biphenylsulfonyl)amino]-3-phenyl-propionic acid is dose-dependent, further supporting the biologic mechanism of action. In our animal model the gelatinolytic activity was greatly increased early on in hepatic venoocclusive disease and this increase in gelatinolytic activity could be attributed to MMP9.<sup>12</sup>

The inventors of the present invention have demonstrated that in the rat monocrotaline model, there is actin depolymerization in sinusoidal endothelial cells and that this in turn leads to increased MMP activity.<sup>13</sup> The causality of the actin depolymerization and increased MMP activity is confirmed by the demonstration that prevention of F-actin depolymerization prevents the monocrotaline-induced increase in matrix metalloproteinase activity. In vitro studies with the various populations of liver cells in vitro have also confirmed that the matrix metalloproteinase activity originates in the sinusoidal endothelial cell rather than in hepatocytes, Kupffer cells or stellate cells.

The following examples are intended to illustrate but not limit the present invention. The methods of the present invention can be further modified for uses such as the identification of drug and diagnostic therapies.

#### EXAMPLE I

The present invention provides methods for using matrix metalloproteinase inhibitors to prevent and treat chemotherapy-induced liver disease, such as SOS and radiation-induced liver disease. These studies were done in an in vivo model of monocrotaline-induced hepatic venoocclusive disease which closely resembles the human disease.<sup>8</sup> Two commercially available MMP inhibitors were tested in the in vivo rat model of hepatic venoocclusive disease: doxycycline and the MMP-2/MMP-9 inhibitor, 2-[(4-biphenylsulfonyl)amino]-3-phenyl-propionic acid. Doxycycline, 15 mg/kg was given twice daily by gavage prior to onset of the disease and continued until the time of sacrifice. 2-[(4-biphenylsulfonyl)amino]-3-phenyl-propionic acid, 200 µg/hour was infused into the portal circulation until the time of sacrifice. A systematic scoring system was devised to review all the changes associated with hepatic venoocclusive disease.

The rats that were treated with the MMP inhibitors were sacrificed on day 4, which is the time-point with most severe disease in this model of hepatic venoocclusive disease when no therapeutic interventions are used. All the livers from the rats treated



with doxycycline or with 2-[(4-biphenylsulfonyl)amino]-3-phenyl-propionic acid were examined blindly by a pathologist according to the scoring system and the pathologist was able to confirm an absence of hepatic venoocclusive disease with doxycycline 15 mg/kg twice daily by gavage or with 2-[(4-biphenylsulfonyl)amino]-3-phenyl-propionic acid, 200 µg/hour infused intraportally by osmotic minipump (see Figure 1). Both doxycycline and 2-[(4-biphenylsulfonyl)amino]-3-phenyl-propionic acid were administered at various doses: lower doses than those mentioned above provided partial protection, whereas the doses listed above completely prevented liver changes in the in vivo model of hepatic venoocclusive disease. Both of these MMP inhibitors prevent hepatic venoocclusive disease in a dose-dependent manner, these showing that this is a class effect of MMP inhibitors and characteristic of MMP inhibition.

In further support of the effect of MMP inhibitors, two analogues of doxycycline were tested for their ability to prevent SOS, anhydrotetracycline and isochlorotetracycline. These two compounds are chemically similar to doxycycline but have little inhibitory effect on MMPs. These analogues had minimal protective effect in the in vivo model (see Figure 1). Furthermore, in the animal model for hepatic venoocclusive disease, the gelatinolytic activity in liver tissue is greatly increased early on and this increase in gelatinolytic activity could be attributed to MMP9. MMP9 mRNA and MMP9 proenzyme also increase very early in the course of disease. No increased gelatinolytic activity could be found in the hepatic vein effluent indicating it is not a non-specific activity in the circulation. In this disease, as MMP9 activity increases, sinusoidal endothelial cell are rounded up resulting in the loss of sinusoidal integrity, which compromises liver microcirculation. Inhibition of the initial rounding up of the sinusoidal endothelial cell by inhibition of matrix metalloproteinases prevents the whole cascade of events.

It will be understood by those skilled in the art that the foregoing illustrates the presently preferred embodiments of the present invention and that modifications may be made in order to accomplish specific ends which do not depart from the spirit of the present invention which is to be limited only by the following claims.

## REFERENCES

1. DeLeve LD, Shulman HM, McDonald GB. Toxic injury to hepatic sinusoids: sinusoidal obstruction syndrome (venoocclusive disease). *Sem.Liver Dis.* 2002;22(1):623-38.
2. Rajvanshi P, Shulman HM, Sievers EL, McDonald GB. Hepatic sinusoidal obstruction following Gemtuzumab Ozogamicin (Mylotarg®). *Blood* 2002;in press.
3. DeLeve LD. Liver Function and Hepatotoxicity in Cancer. In: Holland JF, Frei E, Bast RC, Jr., Kufe DW, Pollock RE, Weichselbaum RR, editors. *Cancer Medicine*. 5th ed. Hamilton, Ontario, Canada: B.C. Decker Inc; 2000. p. Chapter 151.
4. DeLeve LD. Dacarbazine toxicity in murine liver cells: a novel model of hepatic endothelial injury and glutathione defense. *J.Pharmacol.Exp.Ther.* 1994;268:1261-70.
5. DeLeve LD, Wang X, Kuhlenskamp JF, Kaplowitz N. Toxicity of azathioprine and monocrotaline in murine sinusoidal endothelial cells and hepatocytes: the role of glutathione and relevance to hepatic venoocclusive disease. *Hepatology* 1996;23:589-99.
6. DeLeve LD. Cellular target of cyclophosphamide toxicity in the murine liver: role of glutathione and site of metabolic activation. *Hepatology* 1996;24:830-7.
7. DeLeve LD, Ito Y, Machen NW, McCuskey MK, Wang X, McCuskey RS. Embolization by sinusoidal lining cells causes the congestion of hepatic venoocclusive disease. *Gastroenterol.* 2000;118:A2345.
8. DeLeve LD, McCuskey RS, Wang X, Hu L, McCuskey MK, Epstein RB, et al. Characterization of a Reproducible Rat Model of Hepatic Veno-occlusive Disease. *Hepatology* 1999;29:1779-91.
9. Wang X, Kanel GC, DeLeve LD. Support of sinusoidal endothelial cell glutathione prevents hepatic veno-occlusive disease in the rat. *Hepatology* 2000;31:428-34.
10. DeLeve LD, Ito Y, Machen NW, McCuskey MK, McCuskey RS. Sinusoidal dissection and embolization blocks the hepatic microcirculation in hepatic venoocclusive disease. In: *Hepatology*; 1999; 1999. p. 574A.
11. Upadhyaya AG, Harvey RP, Howard TK, Lowell JA, Shenoy S, Strasberg SM. Evidence of a role for matrix metalloproteinases in cold preservation injury of the liver in humans and in the rat. *Hepatology* 1997;26:922-8.
- 12. DeLeve LD, Wang X, Tsai J, Kanel G, Tokes Z. Prevention of Hepatic Venoocclusive Disease in the rat by inhibition of matrix metalloproteinases. *Gastroenterol.* 2001;120:A54.
13. Lamé MW, Jones AD, Wilson DW, Dunston SK, Segall HJ. Protein targets of monocrotaline pyrrole in pulmonary artery endothelial cells. *J.Biol.Chem.* 2000;275(37):29091-9.
14. Read, AE, Weisner RH, LaBrecque DR, et al. Hepatic venoocclusive disease associated with renal transplantation and azathioprine therapy. *Ann Intern Med* 1986;104:651-655.
15. McDonald GB, Hinds MS, Fisher LB, et al. Venocclusive disease of the liver and multiorgan failure after bone marrow transplantation: a cohort study of 355 patients. *Ann Intern Med.* 1993;118:255-267.
16. Jones RJ, Lee KS, Berschoner WE, et al. Veno-occlusive disease of the liver following bone marrow transplantation. *Transplantation* 1987;44:778-783.

17. Carreras E, Bertz H, Arcese W, et al. Incidence and outcome of hepatic veno-occlusive disease after blood or marrow transplantation: a prospective cohort study of the European group for blood and marrow transplantation. *Blood* 1998;92:3599-3604.
18. Shulman HM, McDonald GB, Matthews D, et al. An analysis of hepatic venocclusive disease and centrilobular hepatic degeneration following bone marrow transplantation. *Gastroenterology* 1980;79:1178-1191.
19. Strasser SI, McDonald SJ, Schoch HG, et al. Severe hepatocellular injury after hematopoietic cell transplant: incidence and etiology in 2136 consecutive patients [Abstract]. *Hepatology* 2000;32:299.
20. Kikuchi K, Rudolph R, McDonald GB. Portal vein thrombosis after hematopoietic cell transplant: incidence, treatment, and outcome. *Hepatology* 2000; 32:405 (Abs).
21. Carreras E, Granena A, Navasa M, et al. Transjugular liver biopsy in bone marrow transplantation. *Bone Marrow Transplant* 1993;11:21-26.
22. McDonald GB, Sharma P, Matthews DE, et al. The clinical course of 53 patients with venocclusive disease of the liver after marrow transplantation. *Transplantation* 1985;36:603-608.
23. Wingard JR, Mellits ED, Jones RJ, et al. Association of hepatic veno-occlusive disease with interstitial pneumonitis in bone marrow transplant recipients. *Bone Marrow Transplant* 1989;4:685-689.
24. Zager RA, O'Quigley J, Zager BK, et al. Acute renal failure following bone marrow transplantation; A retrospective study of 272 patients. *Am J Kidney Dis* 1989;13:210-216.
25. Bearman SI, Anderson GL, Mori M, et al. Venocclusive disease of the liver: development of a model for predicting fatal outcome after marrow transplantation. *J Clin Oncol* 1993;11:1729-1736.
26. Bearman SI, Lee JL, Baron AE, et al. Treatment of hepatic venocclusive disease with recombinant human tissue plasminogen activator and heparin in 42 marrow transplant patients. *Blood* 1997;89:1501-1506.
27. Cai T, Fassina G, Morini M, et al. N-acetylcysteine inhibits endothelial cell invasion and angiogenesis. *Lab Invest* 1999;79:1151-1159.
28. Tyagi SC, Ratajska A, Weber KT. Myocardial matrix metalloproteinase(s): localization and activation. *Mol Cell Biochem* 1993;126:49-59.
29. Tyagi SC, Kumar S, Borders S. Reduction-oxidation (redox) state regulation of extracellular matrix metalloproteinases and tissue inhibitors in cardiac normal and transformed fibroblast cells. *J Cell Biochem* 1996;61:139-151.
30. Upadya GA, Strasberg SM. Glutathione, lactobionate, and histidine: cryptic inhibitors of matrix metalloproteinases contained in University of Wisconsin and histidine/tryptophan/ketoglutarate liver preservation solutions. *Hepatology* 200;31:1115-1122.